

SHORT NOTE

New Zealand king shag (*Leucocarbo carunculatus*) with deformed primary feathers

DAVID S. MELVILLE*

1261 Dovedale Road, RD 2 Wakefield, Nelson 7096, New Zealand

ROB SCHUCKARD

4351 Croisilles French Pass Road, RD 3, French Pass 7193, New Zealand

The New Zealand king shag (*Leucocarbo carunculatus*), endemic to the Marlborough Sounds (New Zealand), is 'nationally endangered' (Robertson *et al.* 2017) with a current population estimate of about 800 birds (Schuckard *et al.* 2015; Bell *et al.* 2019). The species has been little studied, in part due to concerns that it was thought to be highly vulnerable to human disturbance (Taylor 2000). Recognising the urgent need to better understand the ecology of this species to inform future management decisions, the Department of Conservation authorised us to capture and band up to four full-grown New Zealand king shags.

We captured one full-grown juvenile New Zealand king shag at Duffers Reef, Marlborough Sounds (40.9562°S, 174.0379°E) on 19 January 2013 using a fishing pole and noose. The bird appeared to be in good condition, but on detailed examination it was found that the outer four primaries on both wings were abnormal (New Zealand king shags have 11 primaries, but the outermost [remic] is much reduced and is not included in this account). The

ventral base of the rachis (the calamus) was swollen and flaky due to what appeared to be deposits of keratin. On the right wing the outer-most primary (P1) was missing, while P3 (numbered ascendantly) was broken near the base (Fig. 1A); on the left wing the outermost primary was broken near the base (Fig. 1B). Apart from dystrophy of the calamus the other affected primaries were of full length, and appeared normal. The other six primaries on each wing also appeared to be in normal condition, as did the rest of the plumage, including the rectrices, which had some wear at the tips (Fig. 1C); there were no fault-bars (Riddle 1908). There was no evidence of mites on the flight feathers from visual inspection. The only other abnormality that we noted were several small lesions on the webs of both feet (Fig. 2).

At the time of capture we had no opportunity to seek veterinary advice or to contact the Department of Conservation as there was no cell phone coverage of the field site.

We collected some samples by taking feather shaft scrape samples of the waxy keratin tissue from the base of the affected primaries. These were submitted to Wildbase, School of Veterinary

Received 29 May 2020; accepted 23 July 2021

*Correspondence: david.melville@xtra.co.nz

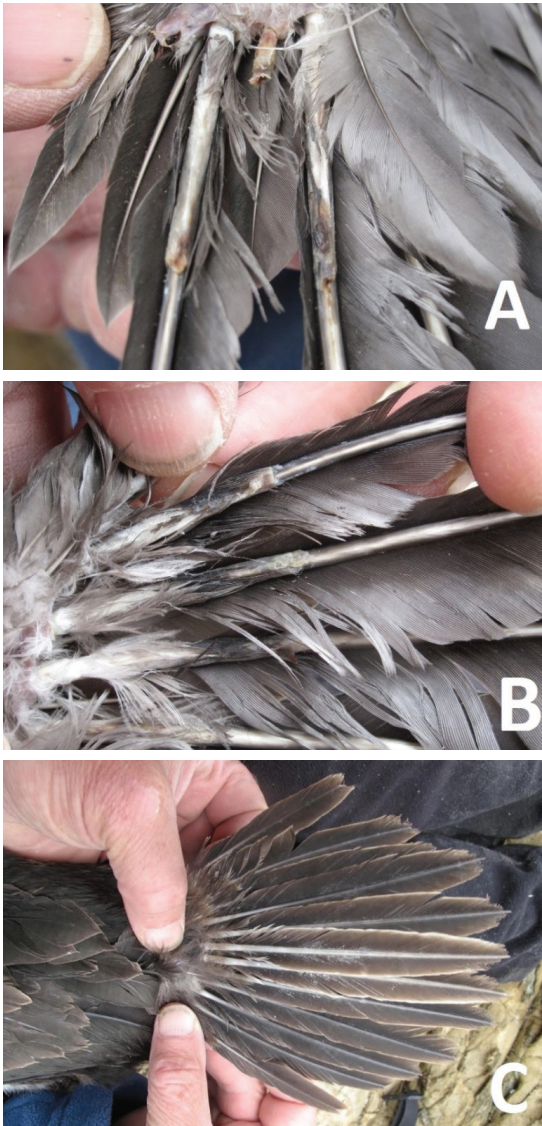


Figure 1. **A** - Outer primaries of right wing of juvenile New Zealand king shag (from below) showing damaged rachis; P1 (outermost primary) missing, P3 broken. **B** - Outer primaries of left wing of juvenile New Zealand king shag (from below) showing damaged rachis; P1 (outermost primary) broken. **(C)** Rectrices of juvenile New Zealand king shag (from above). These feathers did not show any unusual features

Science, Massey University. The results confirmed hyper-keratinisation of the feather shafts, but no cause could be established (Dr Brett Gartrell, *pers. comm.* via email 30 May 2013). Dr Gartrell advised to submit a feather follicle biopsy for histology and culture in future, as plucked or dropped feathers

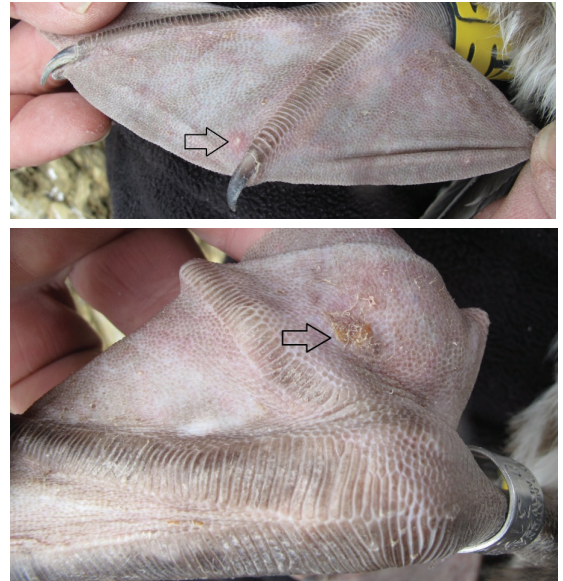


Figure 2. Lesions on the upper surface of the feet of juvenile New Zealand king shag.

are often not diagnostic (Dr Brett Gartrell, *pers. comm.* via email 30 May 2013).

Feather dystrophy, such as that found in our New Zealand king shag, can result from a variety of causes. Psittacine beak and feather disease (PBFD), caused by a circovirus (Todd 2000), can result in feather dystrophy similar that seen in the New Zealand king shag. Whilst primarily recorded in psittacines, PBFD-like symptoms and/or beak and feather disease virus (BFDV) are being recorded in an increasing number of other families and genera of birds (Raidal & Riddoch 1997; Woods & Latimar 2000; Stewart *et al.* 2006; Sarker *et al.* 2015, 2016; Amery-Gale *et al.* 2017). Circoviruses appear to be rarely reported from seabirds, currently only being recorded from three species of gull and two species of penguin (Twentyman 1999; Smyth *et al.* 2006; Morandini *et al.* 2019; Levy *et al.* 2020). The affected primaries appeared to be similar to those affected by 'pinching off syndrome' (POS) (Cooper 1978), which has been recorded in several species of birds of prey in both Europe and North America (Bijlsma & van den Burg 2006; Müller *et al.* 2007a, 2007b; Nemeth *et al.* 2008, 2009; Bijlsma & van de Mortel 2009). The aetiology of POS remains obscure. It was attributed to quill mites (*Harpyrhynchus* spp.) by Heidenreich (1997), while Cooper (2002) and Redig & Cruz-Martinez (2009) suggested that it might be associated with virus infections; it has been found in various North American raptors infected with West Nile Virus (Nemeth *et al.* 2008, 2009).

Müller *et al.* (2007a, 2007b) were unable to attribute POS to any particular cause in European white-tailed sea eagles (*Haliaeetus albicilla*) despite extensive investigation, and concluded that there might be a genetic cause, noting that: 'extremely low genetic drift, possible inbreeding, and the longevity of white-tailed sea eagles may have contributed to the persistence of this disorder' (Müller *et al.* 2007b).

The New Zealand king shag was formerly widespread around the southern coast of the North Island and the northern coast of the South Island (Rawlence *et al.* 2017), but it appears that the population has been restricted to the Marlborough Sounds 'though not in plenty' (Latham 1785), at least since the first specimen was collected in 1773 (Medway 1987). As such, the population may have been subject to a genetic bottleneck, as has been reported in the Stewart Island shag *Leucocarbo chalconotus* (Rawlence *et al.* 2015). If POS is associated with a genetic condition, then it may be found in other individuals within the population.

It seems unlikely that the bird we captured would have been capable of flight, although it did use its wings to help jump from the sea to the rock platform at Duffers Reef. Most New Zealand king shags fly up to 24 km from the colony each day to feed (Schuckard 1994, 2006; Bell 2020); swimming between a colony and a foraging area has been recorded but is uncommon (Bell 2019). Juvenile flight feathers would not normally be replaced until the first complete moult, which Falla (1933) suggests is at about 15 months of age, so it is expected that any juvenile suffering from feather dystrophy would have reduced survival.

Since this initial capture a further 152 New Zealand king shags have been caught and banded (28 adults, 113 chicks, 11 juveniles approximately 2–5 months post fledging) including birds at Duffers Reef. None has shown any evidence of feather dystrophy (Mike Bell *in litt.* 12 July 2021). Vigilance would, however, be advisable. Should any further case of a New Zealand king shag with a plumage disorder be discovered, it is recommended that the live bird be removed from the wild and submitted to an appropriate veterinary facility for detailed examination – future research authorisations should include the ability for such actions.

ACKNOWLEDGEMENTS

This work was undertaken on contract to the Department of Conservation (Contract 4405). We thank Dr Brett Gartrell, School of Veterinary Science, Massey University for examining feather scraping material, and Mike Bell, Toroa Consulting, for providing details of recent New Zealand king shag captures. Janelle Ward and an anonymous referee provided comments which greatly improved the manuscript.

LITERATURE CITED

- Amery-Gale, J.; Marendra, M.S.; Owens, J.; Eden, P.A.; Browning, G.F.; Devlin, J.M. 2017. A high prevalence of beak and feather disease virus in non-psittacine Australian birds. *Journal of Medical Microbiology* 66: 1005–1013.
- Bell, M. 2019. King Shag research project: Year One update report. Unpublished Wildlife Management International Technical Report to the Marine Farming Association and Seafood Innovations Limited. https://www.marinefarming.co.nz/site_files/24792/upload_files/WMIL-KingShagResearchProject-YearOneReport.pdf?dl=1 Downloaded: 22 July 2021.
- Bell, M. 2020. New Zealand King Shag research project: Year Two update report. Unpublished Toroa Consulting Technical Report to the Marine Farming Association and Seafood Innovations Limited. https://www.marinefarming.co.nz/site_files/24792/upload_files/SILMFA-KingShagResearchProject-YearTwoReport.pdf?dl=1 Downloaded: 22 July 2021.
- Bell, M.; Frost, P.; Taylor, G.; Melville, D. 2019. Population assessment during the nonbreeding season of King Shag in the Marlborough Sounds; January 2019. Unpublished Technical Report to New Zealand King Salmon. <https://198i9o1t5qhfqwhf2z86x4y1-wpengine.netdna-ssl.com/wp-content/uploads/2019/07/2019-King-Shag-non-breeding-population-survey-report.pdf> Downloaded: 22 July 2021.
- Bijlsma, R.G.; van den Burg, A. 2006. Veerawijkingen bij nestjonge roofvogels. *De Takkeling* 14: 194–198.
- Bijlsma, R.G.; van de Mortel, T. 2009. Opnieuw veeruitstoot bij een Wespandief *Pernis apivorus*. *De Takkeling* 17: 106–108.
- Cooper, J.E. 1978. *Veterinary aspects of captive birds of prey*. Saul, Gloucestershire, UK, Standfast Press.
- Cooper, J.E. 2002. *Birds of prey: health and disease*. Oxford, Blackwell.
- Falla, R.A. 1933. The King Shag of Queen Charlotte Sounds *Phalacrocorax carrunculatus* (Gmelin). *Emu* 33: 44–48.
- Heidenreich, M. 1997. *Birds of prey: medicine and management*. Oxford, Blackwell.
- Latham, J.D. 1785. *A general synopsis of birds*. Vol. III, Pt. 2. London, Leigh & Sothorby.
- Levy, H.; Fiddaman, S.R.; Djurhuus, A.; Black, C.E.; Krabberger, S.; Smith, A.L.; Hart, T.; Varsani, A. 2020. Identification of circovirus genome in a Chinstrap Penguin (*Pygoscelis antarcticus*) and Adélie Penguin (*Pygoscelis adeliae*) on the Antarctic Peninsula. *Viruses* 12: 858. DOI: 10.3390/v12080858
- Medway, D.G. 1987. King Shags – a correction. *Notornis* 34: 80.

- Morandini, V.; Dugger, K.M.; Ballard, G.; Elrod, M.; Schmidt, A.; Ruoppolo, V.; Lescroël, A.; Jongsomjit, D.; Massaro, M.; Pennycook, J.; Kooyman, G.L.; Schmidlin, K.; Kraberger, S.; Ainley, D.G.; Varsani, A. 2019. Identification of a novel Adélie Penguin circovirus at Cape Crozier (Ross Island, Antarctica). *Viruses* 11: 1088. DOI: 10.3390/v11121088
- Müller, K.; Schettler, E.; Gerlach, H.; Brunnberg, L.; Hafez, H.M.; Hattermann, K.; Johne, R.; Kollmann, R.; Krone, O.; Lierz, M.; Linke, S.; Lueschow, D.; Mankertz, A.; Müller, H.; Prusas, C.; Raue, R.; Soike, D.; Speck, S.; Wolf, P.; Frölich, K. 2007a. Investigations on the aetiology of pinching off syndrome in four white-tailed sea eagles (*Haliaeetus albicilla*) from Germany. *Avian Pathology* 36: 235–243.
- Müller, K.; Altenkamp, A.; Brunnberg, L.; Fašungová, L.; Freymann, H.; Frölich, K.; Kollmann, R.; Krone, O.; Literák, I.; Mizera, T.; Sömmmer, P.; Schettler, E. 2007b. Pinching off syndrome in free-ranging White-tailed Sea Eagles (*Haliaeetus albicilla*) in Europe: frequency and geographic distribution of a generalized feather abnormality. *Journal of Avian Medicine and Surgery* 21: 103–109.
- Nemeth, N.M.; Kratz, G.E.; Bates, R.; Scherpelz, J.A.; Bowen, R.A.; Komar, N. 2008. Naturally induced humoral immunity to West Nile Virus infection in raptors. *EcoHealth* 5: 298–304.
- Nemeth, N.M.; Kratz, G.E.; Bates, R.; Scherpelz, J.A.; Bowen, R.A.; Komar, N. 2009. Clinical evaluation and outcomes of naturally acquired West Nile Virus infection in raptors. *Journal of Zoo and Wildlife Medicine* 40: 51–63.
- Raidal, S.R.; Riddoch, P.A. 1997. A feather disease in Senegal Doves (*Streptopelia senegalensis*) morphologically similar to psittacine beak and feather disease. *Avian Pathology* 26: 829–836.
- Rawlence, N.J.; Kennedy, M.; Anderson, C.N.K.; Prost, S.; Till, C.E.; Smith, I.W.G.; Scofield, R.P.; Tennyson, A.J.D.; Hamell, J.; Lalas, C.; Matisoo-Smith, E.A.; Waters, J.M. 2015. Geographically contrasting biodiversity reductions in a widespread New Zealand seabird. *Molecular Ecology* 24: 4605–4616.
- Rawlence, N.J.; Till, C.E.; Easton, L.J.; Spencer, H.G.; Schuckard, R.; Melville, D.S.; Scofield, R.P.; Tennyson, A.J.D.; Rayner, M.J.; Waters, J.M.; Kennedy, M. 2017. Speciation, range contraction and extinction in the endemic New Zealand King Shag complex. *Molecular Phylogenetics and Evolution* 115: 197–209.
- Redig, P.T.; Cruz-Martinez, L. 2009. Raptors. Pp. 209–242 In: Tully, T.; Dorresteijn, G.; Jones, A. (eds.) *Handbook of avian medicine*. Second edition. England, Saunders Elsevier.
- Riddle, O. 1908. The genesis of fault-bars in feathers and the cause of alternation of light and dark fundamental bars. *Biological Bulletin* 14: 328–371.
- Robertson, H.A.; Baird, K.; Dowding, J.E.; Elliott, G.P.; Hitchmough, R.A.; Miskelly, C.M.; McArthur, N.; O'Donnell, C.F.J.; Sagar, P.M.; Scofield, R.P.; Taylor, G.A. 2017. Conservation status of New Zealand birds, 2016. *New Zealand Threat Classification Series* 19. Wellington, Department of Conservation. 23 pp.
- Sarker, S.; Moylan, K.G.; Ghorashi, S.A.; Forwood, J.K.; Peters, A.; Raidal, S.R. 2015. Evidence of a deep viral host switch event with beak and feather disease virus infection in rainbow bee-eaters (*Merops ornatus*). *Scientific Reports* 5: 14511. DOI: 10.1038/srep14511
- Sarker, S.; Lloyd, C.; Forwood, J.; Raidal, S.R. 2016. Forensic genetic evidence of beak and feather disease virus infection in a Powerful Owl, *Ninox strenua*. *Emu* 116: 71–74.
- Schuckard, R. 1994. New Zealand King Shag (*Leucocarbo carunculatus*) on Duffers Reef, Marlborough Sounds. *Notornis* 41: 93–108.
- Schuckard, R. 2006. Distribution of New Zealand king shags (*Leucocarbo carunculatus*) foraging from the Trio Is and Stewart I colonies, Marlborough Sounds, New Zealand. *Notornis* 53: 291–296.
- Schuckard, R.; Melville, D.S.; Taylor, G. 2015. Population and breeding census of New Zealand king shag (*Leucocarbo carunculatus*) in 2015. *Notornis* 62: 209–218.
- Smyth, J.A.; Todd, D.; Scott, A.; Beckett, A.; Twentyman, C.M.; Brøjer, C.; Uhlhorn, H.; Gavier-Widen, D. 2006. Identification of circovirus infection in three species of gull. *Veterinary Record* 159: 212–214.
- Stewart, M.E.; Perry, R.; Raidal, S.R. 2006. Identification of a novel circovirus in Australian ravens (*Corvus coronoides*) with feather disease. *Avian Pathology* 35: 86–92.
- Taylor, G.A. 2000. Action plan for seabird conservation in New Zealand. Part A: Threatened seabirds. *Threatened Species Occasional Publication No. 16*. Wellington, Department of Conservation 233 pp.
- Todd, D. 2000. Circoviruses: immunosuppressive threats to avian species: a review. *Avian Pathology* 29: 373–394.
- Twentyman, C.M.; Alley, M.R.; Meers, J.; Cooke, M.M.; Duignan, P.J. 1999. Circovirus-like infection in a southern black-backed gull (*Larus dominicanus*). *Avian Pathology* 28: 513–516.
- Woods, L.W.; Latimer, K.S. 2000. Circovirus infection of nonpsittacine birds. *Journal of Avian Medicine and Surgery* 14: 154–163.

Keywords: New Zealand king shag, *Leucocarbo carunculatus*, deformed primaries, feather dystrophy